ing Spectrophotometer whose wave length and optical density scales had been calibrated.

Contribution from Department of Chemistry Massachusetts Institute of Technology Cambridge, Massachusetts

#### Paper Chromatography of Bile Acids<sup>1</sup>

## By David Kritchevsky and Martha R. Kirk Received May 5, 1952

The separation of bile acids by paper chromatography has been investigated in this Laboratory and two solvent systems which give different, reproducible  $R_t$  values for several acids have been found. The two systems are *n*-propyl alcoholammonia-water 90:2:8 and *n*-propyl alcoholethanolamine-water 90:5:5. Of the two systems, the latter concentrates the moving material into a smaller area and is, therefore, preferable for identification or separation.

Using this solvent mixture, we have been able to achieve separation of various mixtures of these bile acids. Although the  $R_t$  values of dehydrocholic, cholic and norcholic acids are close together, we have been able to separate mixtures of desoxycholic, dehydrocholic and cholic acids, and of desoxycholic, dehydrocholic and norcholic acid. In these experiments we have generally observed two distinct spots of the two acids whose  $R_t$  values are close together; in some cases, however, they merge to give one spot. All experiments were carried out using 50  $\gamma$  of material; mixtures contained 50  $\gamma$ of each component.

For identification of the bile acids, a 15% phosphoric acid spray, slightly different from that originally proposed by Neher and Wettstein,<sup>2</sup> was used. The acids appeared as brown or red spots in white light, or displayed a greenish-yellow or pink fluoresence in ultraviolet light.

The results are given in Tables I and II.

## TABLE I

~			-	
Kf.	VALUES	FOR	BILE	ACIDS

It VILLED FOR DIDL HC3DS						
P-M-W <sup>a</sup> 90:5:5	P-A+W 90:2:8	E-A-W 90:2:8	P-A-W 5:2:3			
0.92	0.74	0.66	0.95			
.65	.47	.65	.89			
.71	.52	. 71	. 94			
.69	. 51	.70	.94			
.92	.68	.75	. 94			
	P-M-W <sup>a</sup> 90:5:5 0.92 .65 .71 .69	$\begin{array}{cccc} \mathbf{P}\text{-}\mathbf{M}\text{-}\mathbf{W}^a & \mathbf{P}\text{-}\mathbf{A}\text{-}\mathbf{W} \\ \mathbf{90:5:5} & \mathbf{90:2:8} \\ 0.92 & 0.74 \\ .65 & .47 \\ .71 & .52 \\ .69 & .51 \end{array}$	$\begin{array}{c cccccc} \mathbf{P}\text{-}\mathbf{M}\text{-}\mathbf{W}^a & \mathbf{P}\text{-}\mathbf{A}\text{-}\mathbf{W} & \mathbf{P}\text{-}\mathbf{A}\text{-}\mathbf{W} \\ \mathbf{90;5:5} & \mathbf{90;2:8} & \mathbf{90;2:8} \\ 0.92 & 0.74 & 0.66 \\ .65 & .47 & .65 \\ .71 & .52 & .71 \\ .69 & .51 & .70 \end{array}$			

 $^{\rm e}$  P, n-propyl alcohol; M, monoethanolamine; A, ammonia; W, water.

#### TABLE II

### SEPARATIONS

Mixture	$R_{\rm f}$ values
Desoxycholic/dehydrocholic/cholic	0.95/0.65/0.72
Desoxycholic/dehydrocholic/norcholic	0.92/0.62/0.73

#### Experimental

The organic solvents were distilled prior to use. All mixtures are by volume as given. Whatman #1 paper was used throughout.

The material to be chromatographed was applied to a spot about 2 cm. in diameter on a  $4 \times 40$ -cm. strip of filter

(1) The work described in this paper was sponsored by the United States Atomic Energy Commission.

States Atomic Energy Commission.
(2) R. Neher and A. Wettstein, Helv. Chim. Acta, 34, 2278 (1951).

paper. Descending chromatography was used and after the solvent front had advanced 25-35 cm. from the origin, the strips were removed from the chromatographic chamber (a 7 × 50-cm. test-tube) and air-dried. Prior to spraying, the strips were dried at 80° for 15 minutes. The spray solution was prepared by mixing 10 parts of 85% phosphoric acid with 25 parts each of water and 95% ethanol. After the papers were sprayed, they were kept at 90° for 20 minutes. Generally, cholic and norcholic acids showed up as red or brick colored spots and occasionally one of the other acids appeared as a red spot. In ultraviolet light (Model SL Mineralight, Ultra-Violet Products, Inc., South Pasadena, California) desoxycholic acid exhibited a pink fluorescence. When larger quantities of these acids were used (100-200  $\gamma$ ) they all gave colored spots in white light as well as appearing more readily in the ultraviolet.

The  $R_f$  values were measured from the foremost point of the origin to the leading edge of the spot. The solvent mixtures which included ammonia tended to give some streaking, whereas with ethanolamine spots about 15 mm. in diameter were obtained.

All  $R_t$  values represent the average of a number of experiments.

Acknowledgment.—The authors wish to thank Dr. J. G. Buchanan for several helpful discussions and Dr. R. M. Lemmon for generous gifts of norcholic and triformylnorcholic acids.

RADIATION LABORATORY UNIVERSITY OF CALIFORNIA BERKELEY 4, CALIFORNIA

## A Redetermination of the Kinetic Constants for the System α-Chymotrypsin-Nicotinyl-L-tryptophanamide<sup>1</sup>

# By H. T. HUANG AND CARL NIEMANN<sup>2</sup>

RECEIVED MARCH 29, 1952

In previous studies of the kinetics of the  $\alpha$ -chymotrypsin catalyzed hydrolysis of simple specific substrates<sup>3-8</sup> the enzyme preparations used were obtained from a single source, *i.e.*, Armour and Co., although it is true **that** care was taken to use preparations of different lot numbers in several of the investigations.<sup>4,6</sup> While it has been possible in one instance<sup>4</sup> to compare the  $K_{\rm S}$  and  $k_{\rm 3}$  values of acetyl L-tyrosinamide obtained in these laboratories with those obtained elsewhere<sup>9-13</sup> with different enzyme preparations the fact that differences in the reaction systems and analytical procedures were also involved in the above comparison suggested the desirability of a comparison in which the source of the enzyme preparation was the only variable.

The Armour preparation used most frequently in our previous investigations bore the lot no. 90402. This preparation had been used at three different concentrations in a total of twenty-eight separate experiments to evaluate the

- (1) Supported in part by a grant from Eli Lilly and Co.
- (2) To whom inquiries regarding this article should be sent.

(3) H. T. Huang and C. Niemann, THIS JOURNAL, 73, 1541 (1951).
(4) D. W. Thomas, R. V. MacAllister and C. Niemann, *ibid.*, 73,

- 1548 (1951).
- (5) R. J. Foster and C. Niemann, *ibid.*, 73, 1552 (1951).

(6) H. T. Huang, R. V. MacAllister, D. W. Thomas and C. Niemann, *ibid.*, **73**, 3231 (1951).

(7) H. J. Shine and C. Niemann, ibid., 74, 97 (1952).

(8) H. T. Huang, R. J. Foster and C. Niemann, *ibid.*, 74, 105 (1952).

(9) S. Kaufman and H. Neurath, Arch. Biochem., 21, 245 (1949).

(10) S. Kaufman and H. Neurath, J. Biol. Chem., 180, 181 (1949).

(11) G. W. Schwert and S. Kaufman, ibid., 180, 517 (1949).

- (12) S. Kaufman and H. Neurath, ibid., 181, 623 (1949).
- (13) H. Neurath and J. A. Gladner, ibid., 188, 407 (1951).